

## Keys to Successful Testing

Quality of sample analysed = Quality of result

### Avoid vein collapse when drawing samples



- Minimise suction on the syringe, and do not draw back too quickly.

### Prevent haemolysis



- Use the largest vein and needle appropriate for blood collection.
- Never use any needle smaller than a 23 gauge size.
- Use minimal alcohol on fur/skin.
- Remove the needle from the syringe before dispensing into the blood tube, unless using a closed vacuum blood collection system.



### Ensure the correct ratio of anticoagulant to blood



- Always use the smallest collection tube needed.
- Fill lithium heparin and EDTA tubes to minimum fill line.
- Fill sodium citrate tubes exactly to the fill line.



### Prevent unwanted clotting



- Do not** hold off the vein for more than a few seconds before venipuncture.
- For feline samples collected from the medial saphenous vein: a vacuum blood collection system instead of a syringe is recommended.

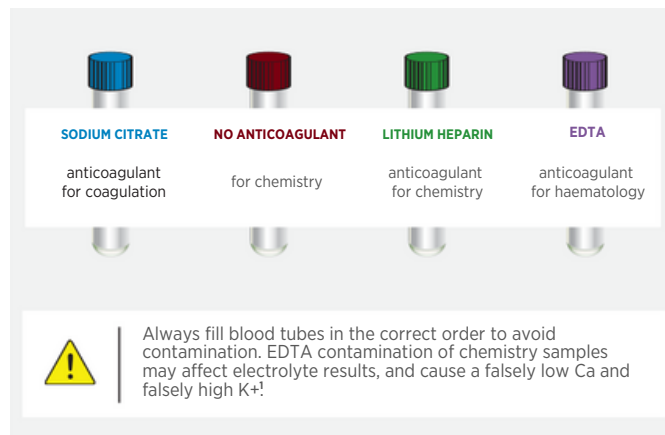


### Do not allow sample to degrade



- Run the sample as soon as possible after drawing.

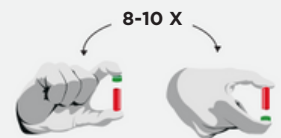
## Collection Tubes & Fill Order



## Tube Handling

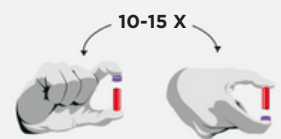
### Chemistry<sup>2,4</sup>

Whole blood samples must be inverted 8-10 times after collection and re-inverted just prior to use.



### Haematology<sup>3,4</sup>

Samples must be inverted 10-15 times after collection and re-inverted just prior to use.



Never shake blood sample tubes.

1.3 mL and smaller tubes may need additional inversions for proper mixing.

Do not rely on a rocker to mix blood samples properly; rockers do not take the place of proper tube inversion.

Zoetis Australia Pty Ltd. ABN 94 156 476 425, Level 6, 5 Rider Boulevard, Rhodes, NSW 2138. © 2022 Zoetis Inc. All rights reserved. March 2024. MM-32960

## Sample Quality



**NORMAL** plasma and serum samples are straw colored, and do not have a yellow, red, or pink tinge.



**HAEMOLYSED** plasma and serum samples have a pink/red tint due to broken red blood cells.

*Avoid haemolysis by using proper sample collection and handling techniques.<sup>1</sup>*



**LIPAEMIC** plasma and serum samples have a milky appearance due to a high concentration of fat in the blood.

*Avoid lipaemia by using a fasted patient sample whenever possible.<sup>1</sup> Remind clients to refrain from feeding their pets prior to their appointment.*



**ICTERIC** plasma and serum samples have a yellow color due to a disease or condition that causes excess bilirubin in the blood.



**CLOTTED** samples may have visible red clots that stick to wooden applicator sticks when swirled in a sample. Traumatic or delayed blood collection can lead to micro and /or macro clots.<sup>1</sup>

*Avoid clotted samples by inverting blood tube appropriately immediately after filling. Re- draw any clotted haematology samples.*

**NOTE:** Never run a clotted sample for analysis on the HM5.

## Sample Storage<sup>5,6</sup>



### Chemistry<sup>2</sup>

Lithium Heparin whole blood samples at room temperature+ must be run within 1 hour,<sup>8</sup> or separated to serum\* or plasma\* and run as soon as possible.<sup>7</sup> Serum and plasma samples may be stored refrigerated ++ for up to 48 hours.<sup>9</sup>



### Haematology<sup>3</sup>

EDTA whole blood samples must be run within 1 hour at room temperature; and may be stored refrigerated++ for up to 12 hours.<sup>7</sup> Blood should return to room temperature prior to running on the HM5.

\* Stored plasma and serum samples must be separated and kept in a stoppered test tube containing no additive.  
+ Room Temperature (20-25 °C)  
++ Refrigerated Temperature (2-8 °C)